



A microfluidic method generating monodispersed microparticles with controllable sizes and mechanical properties

Cheng Zhang^a, Romain Grossier^a, Leda Lacaria^b, Felix Rico^b, Nadine Candoni^a, Stéphane Veessler^{a,*}

^a CNRS, Aix-Marseille Université, CINaM (Centre Interdisciplinaire de Nanosciences de Marseille), Campus de Luminy, Case 913, F-13288 Marseille Cedex 09, France

^b Adhesion and Inflammation Lab (LAI)/INSERM U1067 CNRS UMR 7333, 163, Avenue de Luminy, Case 937, 13288 Marseille Cedex 09, France

HIGHLIGHTS

- Microfluidic method for the production of monodispersed hydrogel microparticles.
- No use of surfactant.
- Microparticle separation is simple and easy.
- Microparticles of controllable sizes and mechanical properties are produced.

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ABSTRACT

Seeking to produce microparticles that mimic red blood cells (RBCs), we present a microfluidic method of generating monodispersed hydrogel microparticles of Na-/Ca-alginate with controllable sizes (micrometer range) and mechanical properties. No surfactant is used. Transformation from Na-alginate to Ca-alginate microparticles is realized via *ex situ* gelation, which proves essential to obtaining desired microparticle properties, such as insolubility in water and RBC-like mechanical properties. For both Na-alginate and Ca-alginate microparticles, a smooth surface and a porous inner structure are observed under a scanning electron microscope. A platform of microgrippers is successfully developed to manipulate the microparticles. The Young's modulus measured using an atomic force microscope on the surface of Ca-alginate microparticles is of the same order as that of RBCs.

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1. Introduction

Microfluidics is recognized as a powerful technique to form discrete and uniform hydrogel structures at the microscale (<100 μm) (Utech et al., 2015). Its main advantage is that droplet-based microfluidic systems permit generation of discrete and uniform microdroplets which are dispersed in a continuous phase (Zhang et al., 2015). Each microdroplet can serve as the site of a gelation reaction that yields microparticles (Hâti et al., 2016; Utech et al., 2015; Yamada et al., 2012; Zhang et al., 2007). In this paper, we explore the generation of monodispersed hydrogel microparticles for potential applications in medical and biological fields (Lee and Mooney, 2012).

Alginate is a well-known biopolymer obtained from brown algae. It has been widely applied in drug delivery and tissue engi-

neering due to its biocompatibility, low toxicity, nonimmunogenicity, relatively low cost, and easy gelation with divalent cations such as Ca²⁺ (ionic cross-linking). Alginate is a linear polysaccharide containing varying amounts of 1,4-linked β-D-Mannuronic acid and α-L-Guluronic acid residues arranged in a block-wise manner along the backbone. The composition and residue sequence vary with the source (Lee and Yuk, 2007). Seeking to control the mechanical and/or swelling properties of alginate gels, researchers have investigated ionic cross-linking with different types of molecules and cross-linking densities (Ouwerx et al., 1998). The purpose of our study was to produce calibrated and monodispersed microparticles whose size (5–10 μm) and mechanical properties (E = 26 ± 7 kPa (Dulińska et al., 2006)) are similar to those of red blood cells (RBCs). This was intended to validate a cellular ultrasonic imaging setup (Monchy et al., 2018) used to assess RBC aggregation under *in vivo* and *in situ* flow conditions.

Calcium alginate (Ca-alginate) was chosen for its mechanical properties (Ouwerx et al., 1998). Alginate forms hydrogel by simple gelation with divalent cations, such as calcium cation (Ca²⁺). However, direct mixing of a calcium solution with an alginate

* Corresponding author at: CNRS, Aix-Marseille Université, CINaM (Centre Interdisciplinaire de Nanosciences de Marseille), Campus de Luminy, Case 913, F-13288 Marseille Cedex 09, France.

E-mail address: veessler@cinam.univ-mrs.fr (S. Veessler).

solution in a microfluidic chip gives rise to instant gelation, likely resulting in clogging and non-uniform microdroplet formation (Utech et al., 2015). To overcome this, other approaches were proposed in the literature to provide calcium (a cross-linker):

(1) dissolution of solid particles: internal gelation when solid particles of calcium carbonate are in the dispersed alginate phase, or external gelation when solid particles of calcium acetate are in the continuous phase (Zhang et al., 2007);

(2) chelation of calcium and its controlled release in the microdroplet via acidification (Utech et al., 2015) or exchange of ligands (Hâti et al., 2016);

(3) injection of a buffer to delay gelation, acting as a barrier to delay calcium diffusion (Yamada et al., 2012) but requiring a complicated microfluidic chip with up to 7 inlet channels and with a risk of clogging.

Thus, *in situ* gelation under a microfluidic method remains an issue. Seeking to overcome this problem, we use a solvent extraction method (Freitas et al., 2005; Rondeau and Cooper-White, 2008; Sugaya et al., 2013). We develop a simple 3-step alginate microparticle synthesis. First, sodium alginate (Na-alginate) microdroplets of the desired size and concentration are produced without using surfactant, since surfactant has been shown to affect particle morphology (Pittermannová et al., 2016). Second, Na-alginate microparticles are obtained by microdroplet shrinkage via water extraction by dimethyl carbonate (DMC). Third, *ex situ* cross-linking is performed with a calcium chloride (CaCl_2) solution, which yields Ca-alginate microparticles. Here, unlike most approaches where particle size is controlled exclusively by channel size (Utech et al., 2015), the final size is controlled by channel diameter, chemical composition of the continuous phase, initial Na-alginate concentration (C_i), ratio (k) of flow rates between the continuous phase and the dispersed phase and total flow rate (Φ_{TOT}) of the two phases.

2. Materials and methods

2.1. Chemical materials

Sodium alginate (I3G80), dimethyl carbonate (ReagentPlus 99%, CAS number 613-38-6,) and calcium chloride (22 317.297) were obtained from Alliance Gums and Industries, SIGMA-ALDRICH and PROLABO, respectively and used without further purification.

2.2. Principle of droplet-based microfluidic system with DMC

A droplet-based microfluidic system is constructed (Fig. 1) using commercially available polyether ether ketone (PEEK) junctions and Teflon-like tubing (IDEX Health and Science) with inner diameter (ID) 150 μm unless specified, initially designed for high-performance liquid chromatography (HPLC) systems. These two materials were previously tested in our group and proved to be compatible with almost all solvents, showing excellent resistance and no solvent evaporation (Ildefonso et al., 2012).

This droplet-based microfluidic system is used to generate hydrogel microparticles dispersed in a flow of continuous phase.

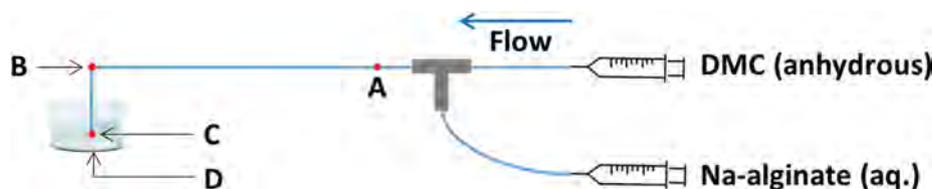


Fig. 1. Diagram of the microfluidic setup where A, B, C and D are observation points after microdroplet generation, at the end of the channel, at the outlet of the channel and at the bottom of the collecting vial respectively.

An aqueous Na-alginate solution (concentration varying over 3 orders of magnitude from 0.006 to 1 wt%) is used as the dispersed phase and dimethyl carbonate (DMC) as the continuous phase. Although these two phases are immiscible at room temperature, water's low solubility (3 wt%) in DMC (Stephenson and Stuart, 1986) ensures slow exchange between DMC and water.

2.3. Principle of shrinkage during generation and collection of microdroplets

Our approach is inspired by Rondeau and Cooper-White (Rondeau and Cooper-White, 2008) who studied the formation of biopolymer microparticles using DMC as continuous phase. Along the microfluidic channel, water contained in Na-alginate microdroplets diffuses slowly into carrier fluid DMC (Fig. 2). Consequently, the concentration of alginate in the microdroplets increases, assuming negligible diffusion of alginate in DMC (Rondeau and Cooper-White, 2008). Furthermore, this mechanism allows the desired microparticle size and concentration to be obtained without using low ID channels or injecting a highly concentrated and viscous alginate solution, both of which would complicate the procedure and risk blocking microchannels. Finally, shrunk microdroplets are collected in a DMC-containing vial, where water extraction continues. Because of the water extraction by DMC, Na-alginate microdroplets generated in the microfluidic channel shrink, becoming Na-alginate microparticles. After gentle drying at room temperature, alginate cross-linking is realized simply by adding an aqueous CaCl_2 solution, forming Ca-alginate microparticles. Fig. 3 shows optical micrographs of microdroplets/particles at different observation points during the experiment. Note that separating Na-alginate and Ca-alginate microparticles from solution is a crucial step for potential applications (Pittermannová et al., 2016).

2.4. Characterization of microparticles

2.4.1. Scanning electron microscope

The topology and inner structure of microparticles are observed by scanning electron microscope (SEM, JEOL 6340F), with a low acceleration voltage of 1 kV and without coating. After drying at room temperature in air, microparticles are fixed to the bottom of a recipient. They are then transferred to a copper sample plot with carbon tape. This step may break some microparticles, revealing the inner structure. A razor blade is also used to cut microparticles to observe the inner structure.

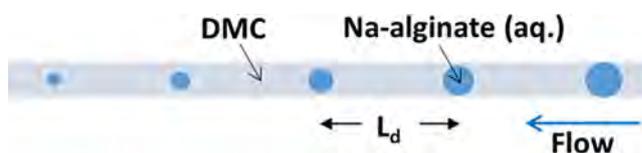


Fig. 2. Schematic illustration of the shrinkage of microdroplets in DMC along the microfluidic channel where L_d is the distance between 2 successive microdroplets.

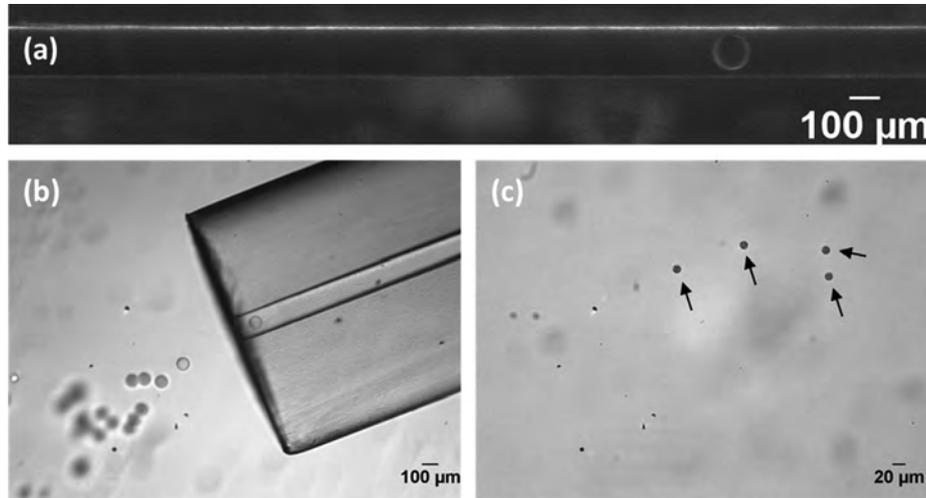


Fig. 3. Optical micrographs of Na-alginate microdroplets ($C_i = 0.06$ wt%) in DMC in a channel (ID 150 μm) (a) after generation (point A in Fig. 1) and (b) at the outlet of the channel (point C in Fig. 1) during collection in DMC. (c) Na-alginate microparticles ($d = 11.7 \mu\text{m} \pm 1.9\%$) in DMC, after complete shrinkage, at the bottom of the collecting vial (point D in Fig. 1).

2.4.2. Microgrippers

The mechanical properties of microparticles are first evaluated using microgrippers (FT-G 102 Force Sensing Microgrippers from FemtoTools). Alginate microparticles, placed in a glass petri dish filled with aqueous solution, are observed under a transmission optical microscope.

2.4.3. Atomic force microscope

The mechanical properties of microparticles are also characterized by measuring the Young's modulus using a NanoWizard atomic force microscope (JPK, Bruker, Santa Barbara, CA, USA). The experiment involves the use of a V-shaped cantilever, an MLCT-Bio-DC(C) (Bruker, Santa Barbara, CA, USA) with a pyramidal probe and a spring constant of 0.01 N/m. Microparticles are deposited on a surface and observed via an inverted optical microscope coupled with AFM.

3. Results and discussion

3.1. In-channel microdroplet shrinkage

We discuss here in detail the shrinkage of spherical microdroplets in the microfluidic channel. For each microdroplet, water diffuses both left and right in the channel (Fig. 4 (a)). To facilitate the calculation, we consider that water diffuses exclusively in one direction, either left or right (Fig. 4 (b)), until DMC is saturated.

Thus, the quantity of DMC in the volume between two microdroplets (V_d), related to the distance between two microdroplets (L_d), determines the maximum quantity of water that can be extracted from one microdroplet. For a microfluidic channel of inner diameter d , V_d is calculated with the Eq. (1).

$$V_d = L_d \times \pi \left(\frac{d}{2} \right)^2 \quad (1)$$

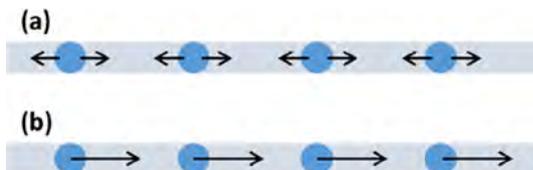


Fig. 4. Illustration of water diffusion (a) in reality and (b) for simplified calculation during extraction by DMC in the channel.

Obviously, given the limited solubility (3 wt%) of water in DMC, a certain L_d (or V_d) is required to achieve the desired decrease in diameter (or volume) of microdroplets.

We demonstrate here a method of estimating the required L_d to obtain a given reduction in diameter after shrinkage of microdroplets of initial diameter d_i . Because diffusion of Na-alginate in DMC is negligible (Rondeau and Cooper-White, 2008), the decrease in microdroplet diameter (or volume) is only due to water extraction by DMC. The desired diameter is attained when DMC is saturated by water. This equilibrium state is described by Eq. (2), which indicates that the volume of water diffused to DMC determines the reduction in microdroplet volume $R_v(\%)$:

$$V_d \times 3\% = \left(\frac{4}{3} \right) \pi \left(\frac{d_i}{2} \right)^3 \times R_v(\%) \quad (2)$$

As a first approximation, we assume that the initial diameter d_i of the microdroplets generated is close to the channel ID ($d_i \approx d$). Then, by combining Eqs. (1) and (2), we get Eq. (3) showing the ratio between required microdroplet distance L_d and initial microdroplet diameter d_i .

$$L_d/d_i \approx \frac{2}{9} R_v \quad (3)$$

As an example, there is a 90% decrease in diameter, from microdroplets with a diameter of 150 μm to microparticles with the same diameter range as RBCs (e.g. 15 μm). This corresponds to a 99.97% decrease in volume R_v . Therefore, according to the theoretical calculation (3), the L_d/d_i ratio required is:

$$L_d/d_i \approx 22.2 \quad (4)$$

3.2. Operating parameters required to obtain desired microdroplet diameter

The channel containing microdroplets and DMC is observed at points A and B, situated 5 cm and 63 cm respectively from the outlet of T-junction (Fig. 1).

3.2.1. Controlling the distance between microdroplets L_d

Experiments are conducted to study the generation of microdroplets at point A, by varying the flow rates of dispersed phase Φ_D and continuous phase Φ_C (Fig. S1 in the supplementary material). Φ_D is varied from 0.001 to 0.3 $\mu\text{L/s}$ and Φ_C from 0.01 to

0.6 $\mu\text{L/s}$. For each pair (ϕ_D and the corresponding ϕ_C), L_d is measured at point **A**. Each pair corresponds to a given ratio k (ϕ_C/ϕ_D). Regarding the total flow rate ϕ_{TOT} ($\phi_D + \phi_C$), our pairs are divided into two groups with low and high values of ϕ_{TOT} . Plotting L_d/d_i versus k for each range of ϕ_{TOT} (Fig. S2 in the supplementary material) shows an increase in L_d/d_i with k , with a linear relationship and slope depending on ϕ_{TOT} . This result is consistent with our previous findings on the influence of ϕ_D/ϕ_C (i.e. $1/k$) and ϕ_{TOT} on microdroplet diameter d_i (Zhang et al., 2015).

Finally, the fitting equation obtained for each value of ϕ_{TOT} gives the value of k required to reach $L_d/d_i = 22.2$ (Eq. (4)), corresponding to a volume reduction of 99.97% (crosses in Fig. S2 in the supplementary material). This means that with a low and a high total flow rate, a value of 21 and 38 for k respectively ensures a volume reduction of 99.97%. Moreover, assuming a maximum reduction of volume $R_v < 100\%$ for the final microparticle, Eq. (3) demonstrates that this R_v is attainable with $L_d/d_i < 22.3$. This is why the fitting curves are stopped at $L_d/d_i < 22.3$. However, loss of water takes time, and meanwhile the microdroplet is moving forward in the channel. Therefore, the length of channel required to reach the final diameter depends on flow rates, as discussed below.

3.2.2. Influence of residence time on shrinkage rate

For a given k , microdroplets are observed at points **A** and **B**. Fig. 5 shows optical micrographs of microdroplets at point **B** obtained for ϕ_{TOT} varying from 0.1 to 0.2 $\mu\text{L/s}$, thus between the two ϕ_{TOT} ranges described above (see fitting curves). Because of k values (20, 30 and 40), L_d is too large to be photographed for measurement. Nevertheless, microdroplet shrinkage is observed for all three values of k . According to optical micrographs taken at points **A** and **B**, microdroplet volume reduces 17%, 71% and 99% respectively for k of 20, 30 and 40. Thus, microdroplet shrinkage or water loss increases with k . Moreover, the significant shrinkage for $k = 40$ is confirmed by observation of microdroplets at different points along the channel (between **A** and **B**), as shown in Fig. S3 of the supplementary material. The final microparticle diameter is about 10 μm , which is in the same range as RBCs.

However, assuming $L_d/d_i = 22.2$ (Eq. (4)) for R_v of 99.97%, i.e. a diameter reduction of 90% (crosses in Fig. 5), experiments should lead to a diameter reduction of around 50%, 80% and almost 100% for k of 20, 30 and 40, respectively. Obviously, the real

shrinkage rate at point **B** is lower than that calculated. This is probably due to the insufficient residence time τ of microdroplets in the channel between points **A** and **B**, which limits the loss of water. Moreover, τ decreases with ϕ_{TOT} since the distance between **A** and **B** is fixed. Therefore, we can either increase the channel length or continue the shrinkage by collecting microdroplets in DMC.

However, a long channel causes fluid instability, probably because of the pressure drop. For instance, microdroplets at the end of the channel for $k = 40$ and $\phi_{TOT} = 0.205 \mu\text{L/s}$ are observed to approach each other, forming aggregations because no surfactant is used (Fig. S4 in the supplementary material). They flow together towards the outlet, despite the fact that they start out relatively far from each other due to the high value of L_d . Actually, because of shrinkage, when microdroplet diameter becomes too small compared to channel ID, microdroplets can move towards the inner wall of the channel and may be affected by contamination there. Our experience indicates that a clean channel is key to avoiding the aggregation of microdroplets. Thus, careful cleaning of the channel with ethanol before experiments is highly recommended, especially when a high k is required. Contrastingly, for low values of k , the main risk is coalescence of microdroplets when leaving the channel. This can be prevented by gentle hydrodynamic agitation. Thus, in addition to the careful choice of k , certain operating techniques also play a role in achieving desired microparticle size.

3.3. Shrinkage during collection: From microdroplets to microparticles of Na-alginate

When collected in DMC, Na-alginate microdroplets continue to shrink and eventually form spherical Na-alginate microparticles. For the experimental condition of $C_i = 0.25 \text{ wt\%}$, **DMC flow rate** = 0.15 $\mu\text{L/s}$, $k = 30$ and $L = 20 \text{ cm}$, d_f is measured on a sampling of about 2386 microparticles and monodisperse size distribution is revealed (Fig. 6).

Initial values of microdroplet diameter (d_i) and Na-alginate concentration (C_i) are varied to study their effect on final microparticle diameter (d_f) and concentration (C_f) after collection. For the experiments in Fig. 7, microdroplets of d_i from 110 to 220 μm are generated with C_i of Na-alginate solutions varying from 0.006 to 1 wt%. For each experiment, d_f is measured on a sampling of about 50 microparticles. It is noteworthy that spherical microparticles with

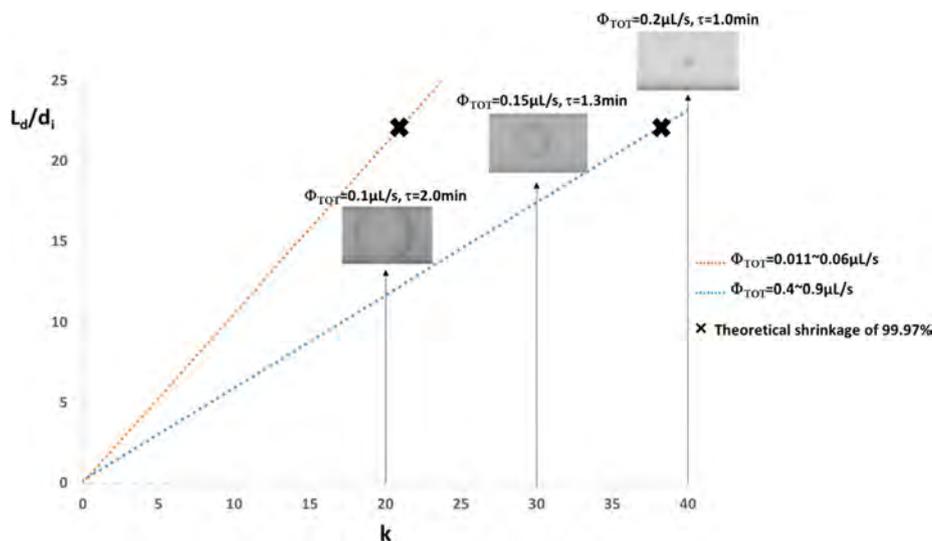


Fig. 5. Fitting of L_d/d_i versus k for two range values of ϕ_{TOT} from Fig. S2 with observations of microdroplets at point **B** (63 cm from T-junction outlet, Fig. 1) for ϕ_{TOT} from 0.1 to 0.2 $\mu\text{L/s}$. Markers (\times) correspond to theoretical shrinkage of 99.97% in microdroplet volume, i.e. 90% diameter reduction ($L_d/d_i = 22.2$). τ : Residence time of microdroplets in the channel. (Initial Na-alginate concentration $C_i = 0.02 \text{ wt\%}$, channel length $L = 70 \text{ cm}$.)

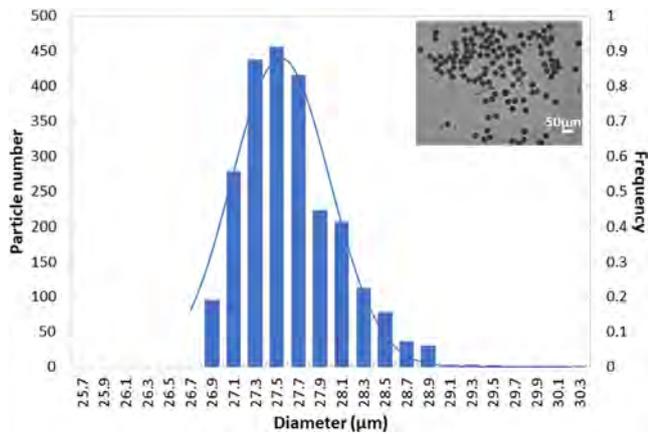


Fig. 6. Size distribution of Na-alginate microparticles. The microparticles are illustrated in the inset. $d_f = 27.5 \mu\text{m} \pm 0.5 \mu\text{m}$. The curve shows the Gaussian fitting. Experimental conditions: $C_i = 0.25 \text{ wt}\%$, **DMC flow rate** = $0.15 \mu\text{L/s}$, $k = 30$ and $L = 20 \text{ cm}$. Result based on 2386 microparticles produced at a frequency of 2 microparticles/s without using surfactant.

a post-collection diameter d_f of $7.7 \mu\text{m} \pm 0.3 \mu\text{m}$ are obtained with $C_i = 0.006 \text{ wt}\%$ (Fig. 10a and b), indicating that RBC size can be reached using this process. Moreover, Fig. 7a shows that d_f increases with both C_i and d_i . Similar results were previously found for porous microparticles of Na-poly(styrenesulfonate) formed by inkjet injection (Yang et al., 2015). In our study, for each C_i , d_f versus d_i is fitted by a power law. According to the fitting equations, the exponent is about 0.3–0.4 for all C_i , whereas the constant factor increases with C_i (Fig. 7a). Since d_f does not greatly vary with d_i when C_i is fixed, a mean value of d_f is calculated for each C_i and plotted versus C_i (Fig. 7b). This leads to another power law dependence. This curve enables the initial concentration of Na-alginate to be defined so as to obtain a given final microparticle size.

In addition, the final concentration of Na-alginate (C_f) in the microparticle after collection is estimated from initial and final volumes of the microdroplet (V_i and V_f), considering that the quantity of Na-alginate in the microdroplet remains constant. Therefore, we assume as above that water diffusion from microdroplets to DMC is the only mass transfer. Then C_f is given by Eq. (5).

$$C_f = C_i \times \left(\frac{d_i}{d_f}\right)^3 \quad (5)$$

From data in Fig. 7a and Eq. (5), C_f is calculated for different d_i and C_i (Fig. 8). It is obvious that C_f depends drastically on d_i , whatever the value of C_i . Thus, a power law fits all the points without

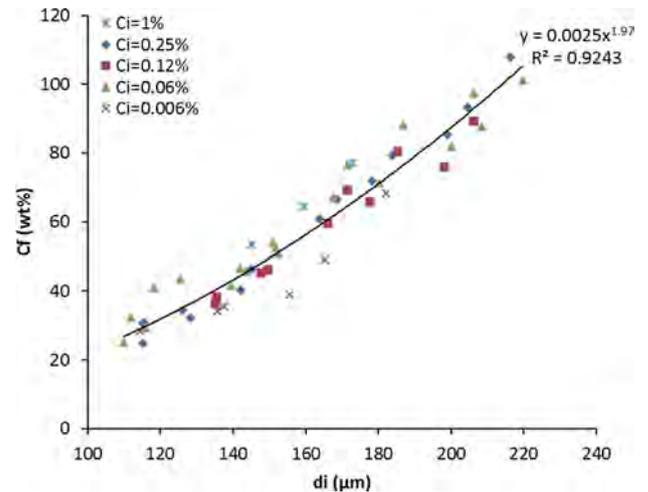


Fig. 8. Relationship between the calculated final Na-alginate concentration C_f in the microparticle and the initial diameter d_i of droplets with different initial concentrations C_i , with the fitting curve of all data.

distinction between C_i values. Note that the fitting equation can be derived from those in Fig. 7a (mathematical derivation in the supplementary material). This curve enables the initial size of droplets to be defined so as to obtain a given final concentration of Na-alginate in microparticles. This final concentration is important, as it can affect the mechanical properties of the microparticles. Thus, the initial size of droplets determines the final concentration of Na-alginate in microparticles, which in turn may control microparticle mechanical properties.

3.4. From Na-alginate microparticles to Ca-alginate microparticles

All the Na-alginate microparticles are observed to be soluble in water, which limits the chemical composition of the media used in further research or applications. To optimize, we therefore use a divalent cation, Ca^{2+} here, for alginate gelation.

3.4.1. Formation of Ca-alginate microparticles by cross-linking with a CaCl_2 solution

After total evaporation of DMC, an aqueous CaCl_2 solution is added to the collecting vial containing dried Na-alginate microparticles. The cation of calcium (Ca^{2+}) induces cross-linking (Yang et al., 2015), thus transforming Na-alginate microparticles into Ca-alginate microparticles. Immediate swelling is observed when CaCl_2 (0.5%, 1% or 10 wt%) is added. The diameter of microparticles

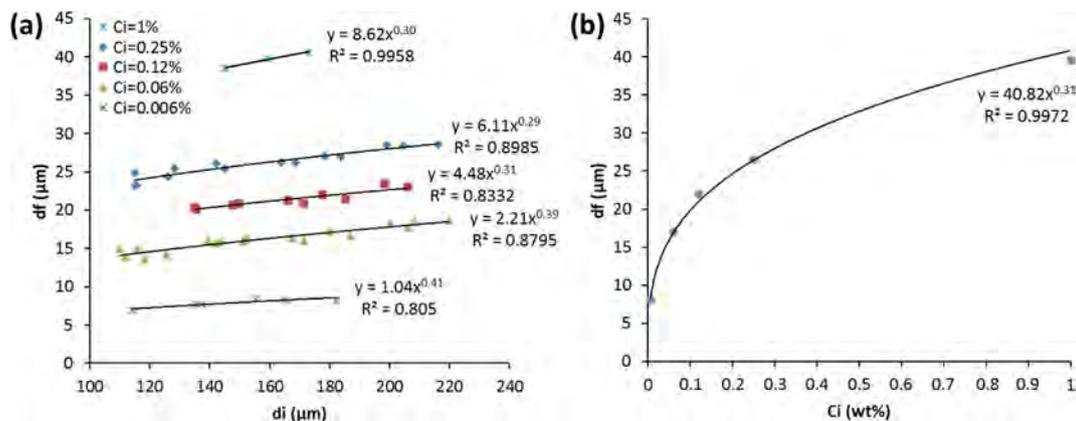


Fig. 7. (a) Final microparticle size d_f as a function of initial microdroplet size d_i for different Na-alginate concentrations C_i (0.006 to 1 wt%); (b) Plot of final microparticle mean diameter d_f after collection versus initial Na-alginate concentration C_i .

increases from $28.7 \mu\text{m} \pm 0.3 \mu\text{m}$ (measured on 51 microparticles) before cross-linking (dried microparticles obtained with $C_1 = 0.25$ - wt%, $k = 5$ and $\phi_{\text{TOT}} = 0.06 \mu\text{L/s}$) to $33.4 \mu\text{m} \pm 0.4 \mu\text{m}$ (measured on 51 microparticles) after immersion and cross-linking (Fig. 9 for 0.5 wt% of CaCl_2). The results are identical whatever the CaCl_2 concentration: it has no obvious effect on either the shape or the size of final Ca-alginate microparticles. Unlike Na-alginate microparticles, all the Ca-alginate microparticles are observed to be insoluble in water. This holds promise for future applications in water.

It is noteworthy that gelation was also achieved for small Na-alginate microparticles ($7.7 \mu\text{m} \pm 0.3 \mu\text{m}$) whose diameter is close to that of RBCs (Figs. 7, 10a and b). Observations lead to the same conclusion: immediate swelling after immersion in CaCl_2 (Fig. 10c). However, due to the difficulty of observation and measurement of the Ca-alginate microparticles produced in this experimental condition, we cannot provide quantitative data regarding size and mechanical properties on these microparticles.

3.4.2. Characterization of Na-alginate and Ca-alginate microparticles

In addition to the optical microscope, the scanning electron microscope (SEM) is also used to observe the topology and the inner structure of microparticles. Furthermore, preliminary attempts to characterize microparticle mechanical properties are carried out using microgrippers.

3.4.2.1. Topology of Na-alginate and Ca-alginate microparticles. SEM images (Fig. 11) show that both Na-alginate and Ca-alginate microparticles are spherical with a smooth surface. Ca-alginate microparticles are slightly smaller than Na-alginate microparticles,

which seems inconsistent with the swelling previously observed (Fig. 9). This can be explained by the manipulating conditions under vacuum in SEM. Because water remaining in the microparticle is extracted and quickly evaporates, there is a second shrinkage after production, until a new equilibrium is reached. This second shrinkage sometimes even causes deformation in the microparticle surface (Fig. S6(b) in the supplementary materials), depending on microparticle properties such as diameter and concentration. This is why, for both Na-alginate and Ca-alginate microparticles, the diameter measured by SEM is smaller than that measured by optical microscope. Moreover, SEM measurements show Na-alginate microparticles to be larger than the corresponding Ca-alginate ones, suggesting that Ca-alginate microparticles lose more water due to the vacuum.

3.4.2.2. Inner structure of Na-alginate and Ca-alginate microparticles. Observations of an Na-alginate microparticle in DMC under optical microscope (Fig. S5 in the supplementary material) and a dried one under SEM (Fig. 12 (a)) reveal a porous inner structure in both cases. We therefore propose the following mechanism of microparticle formation, in accordance with (Yang et al., 2015) First, after Na-alginate microdroplets are generated, water moves from the microdroplets to DMC. Within the microdroplets, this water movement also induces the migration of alginate molecules towards the interface with DMC. As a result, (1) an outer shell of alginate forms gradually until it becomes so dense that water can no longer be extracted and therefore remains trapped inside; (2) several inner areas become vacant after the alginate migrates, which explains the observation of pores. The inner structure of

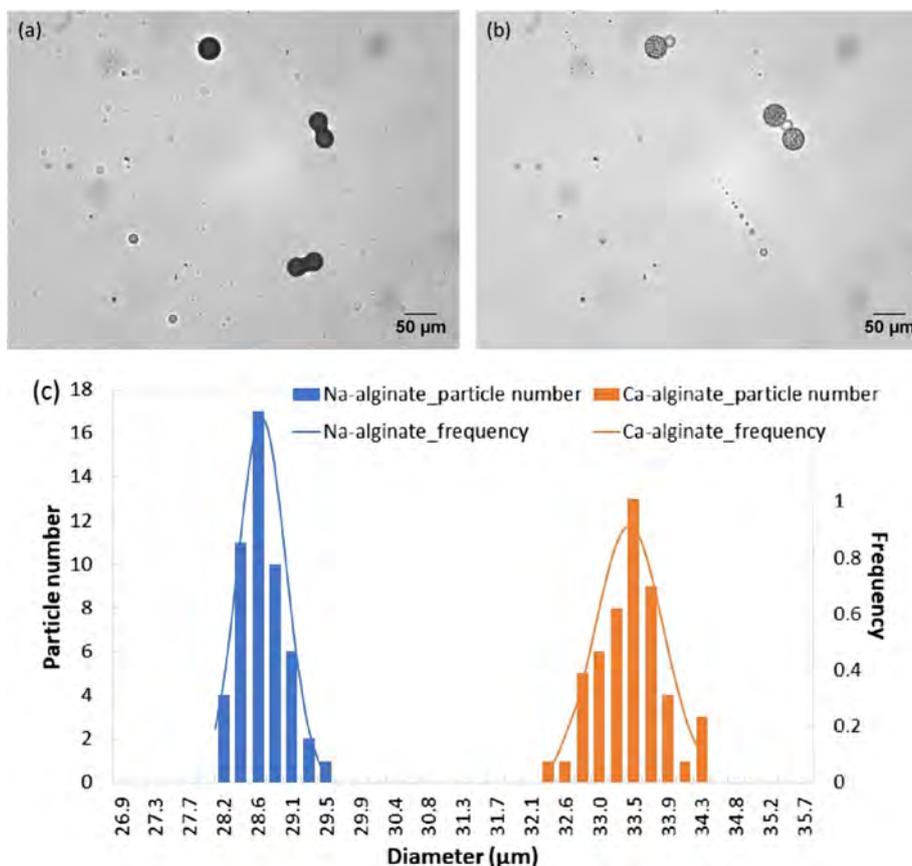


Fig. 9. Images of (a) dried Na-alginate microparticles and (b) Ca-alginate microparticles after immersion in CaCl_2 solution (0.5 wt%). (c) Size distribution of Na-alginate and Ca-alginate microparticles (measured on 51 microparticles). The curves show the Gaussian fitting. Experimental conditions: $C_1 = 0.25$ wt%, DMC flowrate = $0.05 \mu\text{L/s}$ and $k = 5$.

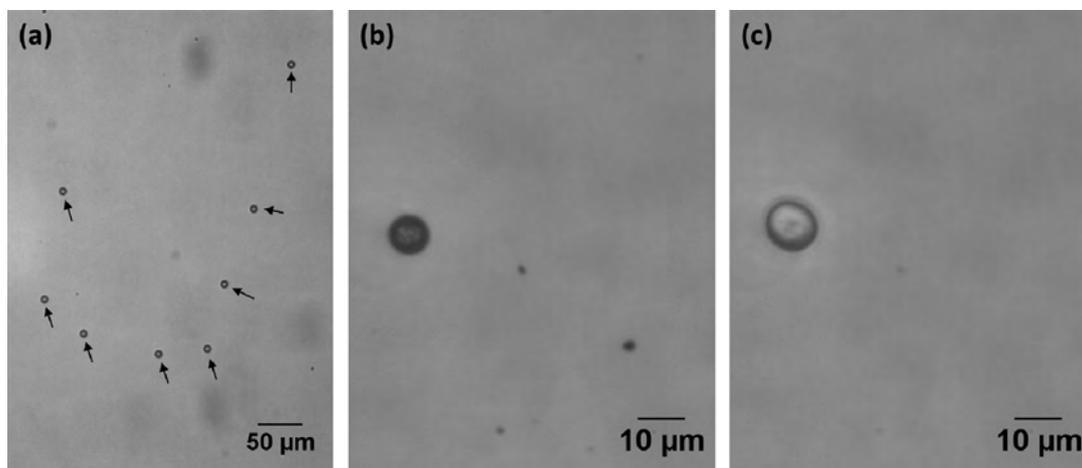


Fig. 10. (a) Alginate microparticles in DMC; (b) a dried alginate microparticle; (c) the same microparticle as in (b) immersed in CaCl_2 (1.35 wt%). Experimental conditions: $C_i = 0.006$ wt%, DMC flow rate = $0.15 \mu\text{L/s}$, $k = 30$, $L = 30$ cm.

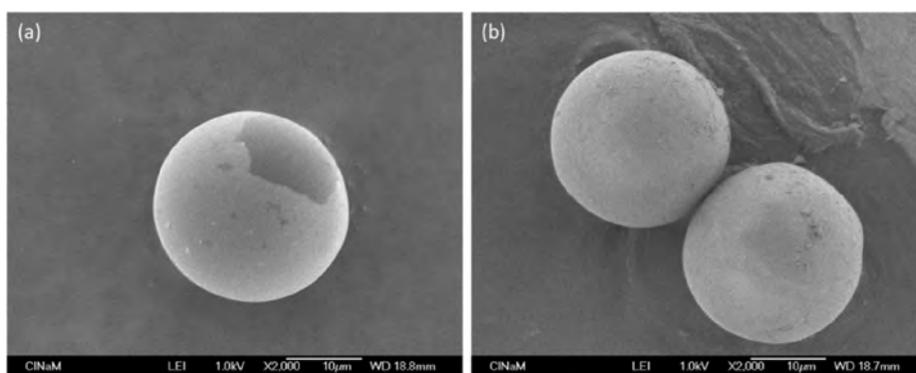


Fig. 11. SEM photographs of (a) Na-alginate and (b) Ca-alginate microparticles (formed in a 0.5 wt% CaCl_2 solution). (Same experimental conditions as for Fig. 9).

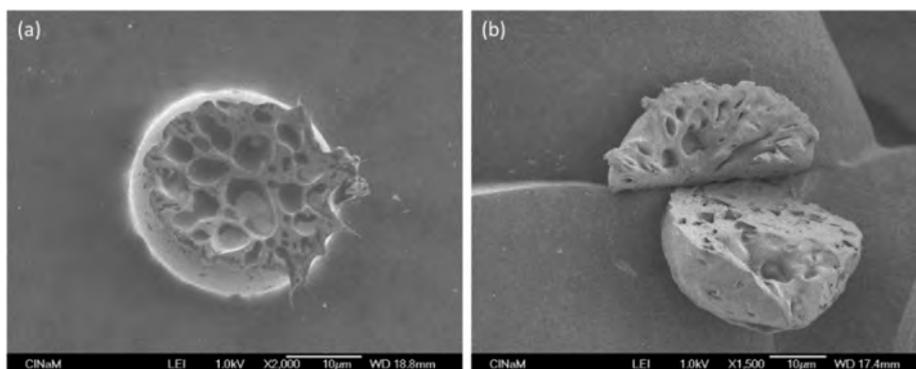


Fig. 12. SEM photographs of the inner structure of (a) Na-alginate and (b) Ca-alginate microparticles (formed in a 0.5 wt% CaCl_2 solution). (Same experimental conditions as for Fig. 9).

Ca-alginate microparticles is also observed to be porous (Fig. 12 (b) and Fig. S7 in the supplementary material), suggesting that cross-linking by calcium cation does not influence microparticle inner structure.

3.4.2.3. Mechanical properties evaluated by microgrippers. After drying, Na-alginate and Ca-alginate microparticles are grasped with force sensing microgrippers to estimate their mechanical properties (Fig. S8 in the supplementary material). Using a 6D positioning

system, the microgrippers are placed around the microparticle to be tested. Depending on the voltage supplied to the microgripper left arm (0–170 V), the microparticle is grasped and the force sensor of the right arm provides applied force of up to $250 \mu\text{N}$. This applied force combined with the optical observation provides access to the microparticle reaction to the compression. For the Na-alginate microparticle (Fig. 13), we observed (Fig. 13 (c)) slipping of the left gripper arm, which indicates a strong resistance to the maximum applied force of $250 \mu\text{N}$. For both Na-alginate

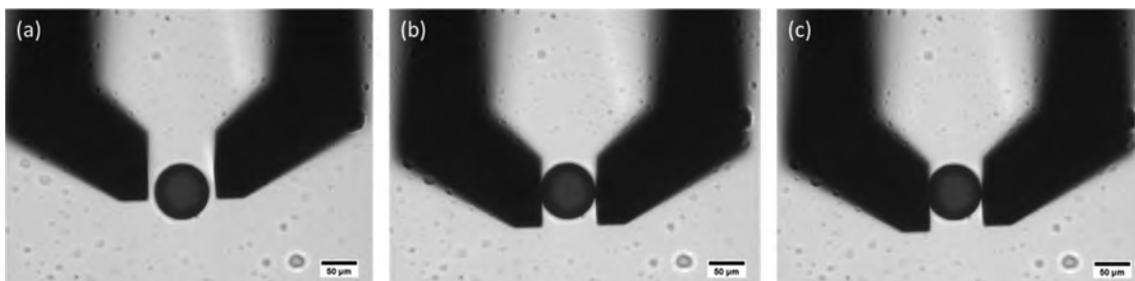


Fig. 13. A dried Na-alginate microparticle ($d_f = 78.7 \mu\text{m}$, $C_i = 0.25 \text{ wt\%}$, **DMC flow rate** = $1.5 \mu\text{L/s}$, $k = 50$, $L = 30 \text{ cm}$ in a capillary with inner diameter (ID) $500 \mu\text{m}$) between microgrippers during (a) approach, (b) contact and (c) force-controlled gripping.

and Ca-alginate microparticles manipulated by microgrippers, no overall deformation was observed.

3.4.2.4. Mechanical properties measured by AFM. The Young's moduli of both Na-alginate and Ca-alginate microparticles are measured by AFM (Fig. 14 (a)). All measurements are performed under the same force of 0.4 nN , an approach velocity of $2 \mu\text{m/s}$ and a z length of $1 \mu\text{m}$. An 8×8 force curve grid (points $2.5 \mu\text{m}$ apart) is acquired in the center of the microparticle (Fig. 14 (b)). Each point is measured 5 times to obtain 5 force curves/point.

An example of a force curve is shown in Fig. 15. The vertical deflection of the cantilever, proportional to the force, depends on the distance between AFM probe and microparticle surface. During measurement, the AFM probe indents about 320 nm on the surface (Fig. 15), less than 1% of the diameter of Ca-alginate microparticles ($35 \mu\text{m}$). Thus, the Young's modulus is measured at the microparticle surface.

Furthermore, we find that the Young's modulus of Na-alginate microparticles is of the order of MPa, whereas the Young's modulus of Ca-alginate microparticles is $47 \pm 43 \text{ kPa}$. Obviously, there is a significant decrease in Young's modulus from Na-alginate microparticles to Ca-alginate microparticles. This is due to the swelling observed during the gelation process when Na-alginate microparticles are immersed in CaCl_2 solution. Consequently, Ca-alginate microparticles contain more water than Na-alginate microparticles, reducing the Young's modulus.

Moreover, the Young's modulus of Ca-alginate microparticles ($47 \pm 43 \text{ kPa}$) is of the same order as that of RBCs ($26 \pm 7 \text{ kPa}$),

where a similar indentation of 200 nm was reported using a similar AFM probe. (Dulińska et al., 2006) It should be noted that larger Young's modulus values have been found in pathological RBCs (e.g. $90 \pm 20 \text{ kPa}$ for erythrocytes with G6PD deficiency) (Dulińska et al., 2006) and in RBCs that have undergone specific treatments (e.g. $191 \pm 28 \text{ kPa}$ for formalin-treated cells) (Girasole et al., 2012). Nevertheless, the Young's modulus of Na-alginate microparticles ($\sim\text{MPa}$) remains beyond the latter range. Gelation can therefore be considered a key step in producing microparticles with mechanical properties close to those of RBCs.

On the other hand, we also find that the Young's modulus varies substantially depending on the measuring point on the microparticle surface (Fig. S9 in the supplementary material). Moreover, this variation also depends on the individual microparticle, even though all are prepared in the same way and come from the same experimental batch. This is actually reflected by the relatively high standard deviation in results ($E = 47 \pm 43 \text{ kPa}$). Surface irregularities can lead to Young's modulus variations; however here, the surface of both Na-alginate and Ca-alginate microparticles is smooth according to SEM photographs (Fig. 11). Thus, it may be reasonable to attribute the variation to the microparticles' porous inner structure (Figs. 12 and 16). As shown in Fig. 16, the thickness of the shell, defined as the space between the surface and the closest inner pore of the microparticle, varies greatly. Because of the heterogeneity of the porous inner structure, this thickness can be of the same order (500 nm) as the indentation of the cantilever probe (320 nm), or larger (in μm range). Consequently, the varying Young's moduli measured may be the result of varying shell thickness. However, further investigations would be required to validate this explanation.

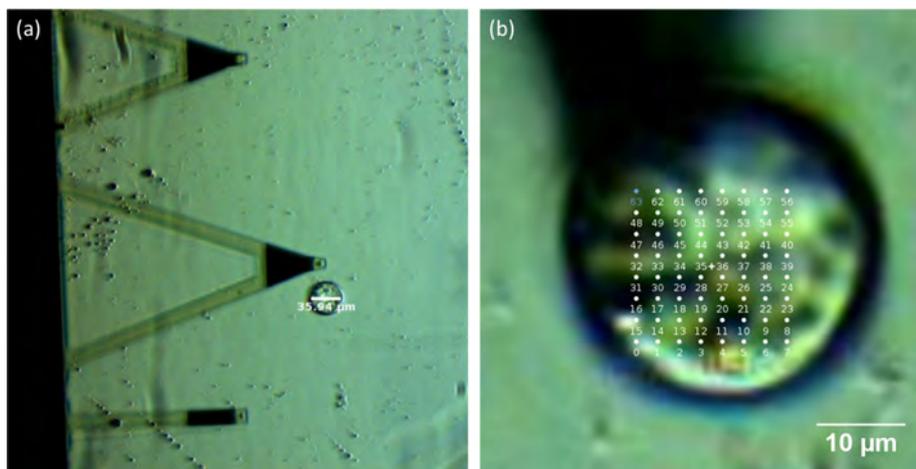


Fig. 14. Image of (a) the cantilever used for the force measurement by AFM and a Ca-alginate microparticle in water ($C_i = 0.25 \text{ wt\%}$, **DMC flow rate** = $0.15 \mu\text{L/s}$, $k = 50$, $L = 19 \text{ cm}$ and $[\text{CaCl}_2] = 0.5 \text{ wt\%}$) and (b) an example of a force curve grid on a Ca-alginate microparticle, acquired during the experiment using an inverted optical microscope coupled with AFM.

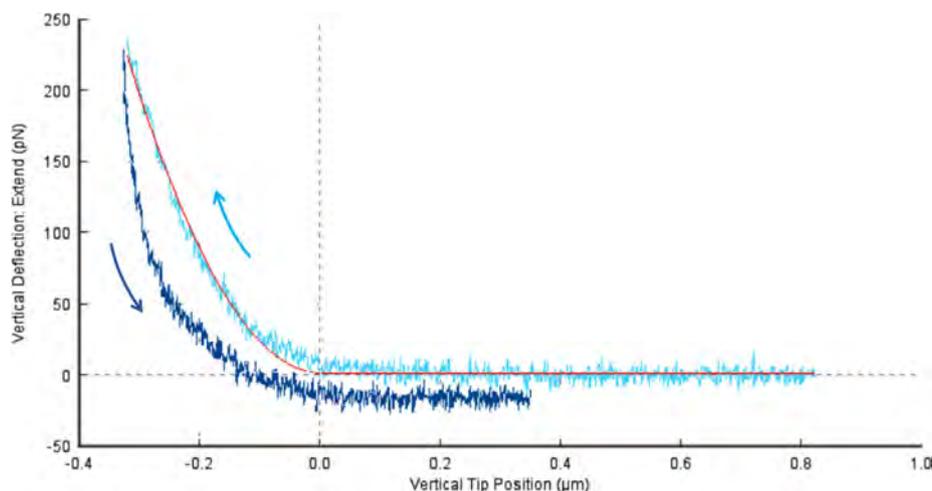


Fig. 15. Example of a force curve on a Ca-alginate microparticle: curves relative to cantilever approach and retraction, indicated by arrows, are respectively in light and dark blue; the Hertz Model fit is in red. Zero vertical position shows the point of contact between cantilever and microparticle where no force is applied and the distance between cantilever and sample surface is zero. The negative values of the abscissa, in absolute value, are the indentation, which is the bead surface deformation. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

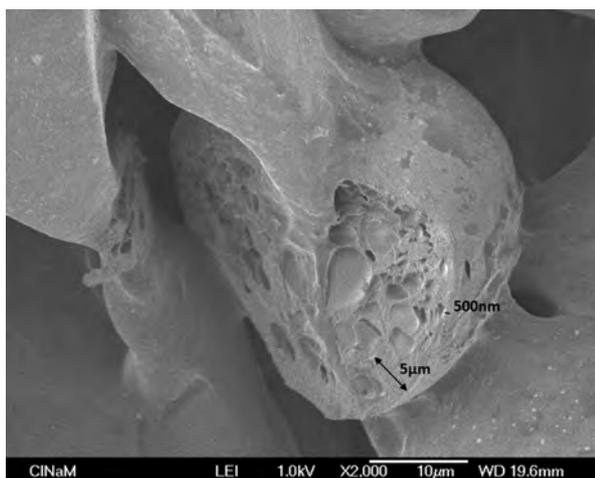


Fig. 16. SEM photograph of the inner structure of a Ca-alginate microparticle.

4. Conclusions and perspectives

In this article, we present a microfluidic method of producing monodispersed hydrogel microparticles of Na-/Ca-alginate with controllable sizes (micrometer range) and mechanical properties without using surfactant. Here, our ultimate aim was to produce microparticles that mimic red blood cells (RBCs). *Ex situ* gelation, used for the transformation from Na-alginate to Ca-alginate microparticles, proves key to obtaining the desired microparticle properties, such as insolubility in water and RBC-like mechanical properties. For both Na-alginate and Ca-alginate microparticles, a smooth surface and a porous inner structure are observed under a scanning electron microscope. A platform of microgrippers was successfully developed to manipulate the microparticles. The Young's modulus, measured using an atomic force microscope on the surface of the Ca-alginate microparticles generated, is of the same order as that of RBCs. Combined with size control to micrometer range, this confirms the feasibility of using Ca-alginate microparticles to mimic RBCs and subsequently study the aggregation of RBCs under *in vivo* and *in situ* flow conditions with ultrasonic imaging. Moreover, we describe a simple method of separating Na-alginate and Ca-alginate microparticles from solution by gentle drying at room temperature.

Interesting future perspectives include studying parameters such as the concentration of CaCl_2 and pH, to determine whether they have an impact on the mechanical properties of Ca-alginate microparticles. This would lead to better understanding and control of microparticle mechanical properties.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ces.2019.115322>.

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